# An updated dichotomous key to the snakes of Europe

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This work presents an updated dichotomous key to all snake species of Europe, based on morphological, morphometric and distributional characters. It gathers data from both the last dichotomous keys concerning European snake species and every following paper giving updates on taxonomic classification or the occurrence of new allochthonous species. The key includes 58 snake taxa, including 57species and one still unnamed taxon, distributed on the European territory, and photographic material for each species. The aim of this paper is to make the identification of all European snake species through a single manuscript more accessible, not only for researchers, but also for citizen science, thus enhancing data collection and assist species conservation.

Key words: dichotomous key; European territory; Ophidia; snakes; species identification.

A dichotomous key is a powerful tool for classification and identification of both objects and above all living organisms (Watson, 2009). It provides the reader with a simple choice method between two paired statements describing alternative morphological characteristics. Once the choice is made, the key guides the user to another couplet of statements, repeating the process until the object or organism is identified (Van Sinh et al., 2017; Armi-ÑANA-GARCÍA et al., 2020). Dichotomous keys are widespread tools among researchers, technicians and students, but can be useful even to people not belonging to the scientific world (DI NICOLA, 2019). This is important to improve the socalled "citizen science", which is becoming very popular both for Europe and single countries (Vohland *et al.*, 2021), in particular when referred to technological and social innovation leading to social change (Butkevičienė *et al.*, 2021).

The scientific literature is increasingly enriched with new taxonomic keys concerning plant and animal species. Snakes are no exception (just in the last few years, it is possible to cite examples as: Di Nicola, 2019; Mallik et al., 2019; Weinell et al., 2019; Alshammari & Busais, 2020; Ganesh et al., 2020; Spengler Waltrick & Menta Giasson, 2021; Dávalos-Martínez et al., 2021).

Over the years, dichotomous keys for snakes present on European or Mediterranean areas have already been created both for popular and scientific literature (e.g. BOULENGER, 1913; VENCHI & SINDACO, 2006;

Speybroeck *et al.*, 2016), but the numerous scientific researches in the herpetological field, beside an increasing use of molecular techniques and with an even better knowledge of the investigated territories, involve continuous taxonomic updates and more in-depth knowledge of the species distribution. Hence the need to have an updated key, in line with recent systematic updates, with the current distributions of the species and with the presence of alien taxa that settled with stable populations in the last years.

#### MATERIALS AND METHODS

The dichotomous key proposed in this work is based on morphological and morphometric characters and on the snake finding locations; it is realized taking as a reference the keys present in the following literature: Boulenger (1913); Venchi & Sindaco (2006); Corti et al. (2011); Sindaco et al. (2013); Speybroeck et al. (2016); Geniez (2018); Di Nicola (2019).

In addition, to obtain updated information at the taxonomic, anatomical (morphological descriptions) and distributional level, the following publications were consulted: Guicking et al. (2006); Kreiner (2007); Cabrera-Pérez et al. (2012); VAN DE KOPPEL et al. (2012); Mahlow et al. (2013); Silva-Rocha et al. (2015); KINDLER (2018); KINDLER & FRITZ (2018); Mizsei et al. (2018); Faraone et al. (2019a, b); Jablonski et al. (2019); Paolino et al. (2019); Zaher et al. (2019); Akbarpour et al. (2020); CATTANEO (2020); CLEMENS & Allain (2020); Freitas et al. (2020); Fritz & Schmidtler (2020); Fritz et al. (2020); Seghetti et al. (2020); Speybroeck et al. (2020); Tamar et al. (2020); Wallach (2020); DI NICOLA *et al.* (2021); VAN DOORN *et al.* (2021).

The European area considered (Fig. S1), is in agreement with what was established by Speybroeck *et al.* (2020), thus including all territories of the European mainland, Macaronesia (except Cape Verde), Balearic Islands, all Greek and Italian islands, Malta and Cyprus. The easternmost limits of the area include territories west of the Ural Mountains and the Ural River (SW Sverdlovsk Oblast, NW Čeljabinsk Oblast and W Kazakhstan included), north of the Caucasus Mountains and NE Azerbaijan.

The key covers species level only, and does not consider subspecies since there are many taxa requiring a status overhaul (e.g., subspecies are sometimes still based on old and not always reliable morphological descriptions). The key also considers allochthonous snakes, but only species that are present in Europe with reproductive populations have been included, such as Elaphe schrenckii, Elaphe taeniura, Indotyphlops braminus and Lampropeltis getula (Cabrera-Pérez et al., 2012; van de Koppel et al., 2012; Mateo, 2013; Faraone et al., 2019a; Paolino et al., 2019; Struijk et al. 2020; Vella et al., 2020; Di Nicola et al., 2021; VAN DOORN et al., 2021). Rhynchocalamus melanocephalus from Cyprus is also considered, due to three different findings in the period 2013-2018 (TAMAR et al., 2020). The allochthonous/recently introduced status was made explicit only for species whose allochtony is recent (with the first records occurring in the last decades) and supported by the literature. For taxa likely introduced but whose origin hypotheses are still unclear (e.g. Eryx jaculus for Sicily and R. melanocephalus for Cyprus) the status in the key has not been specified.

Higher taxa (i.e. superfamilies and families) maintain the same partition explained and provided by ZAHER et al. (2019). Family Viperidae is separated from all other colubroideans (thus explaining why it does not currently belong to any superfamily), within the robustly supported clade Endoglyptodonta. Colubroideans are represented by superfamilies Elapoidea sensu stricto and Colubroidea sensu stricto, which are sister clades with robust support. The former contains family Psammophiidae (here represented by genus Malpolon) with strong support; while the latter contains family Natricidae (robustly supported) as the sister group (no support) of a bigger clade formed by (moderate support) Colubridae, Graviidae, Calamariidae, and Sibynophiidae. Thus, Natricidae and Colubridae are listed as distinct taxa.

Superfamily Booidea is represented by genus *Eryx*, which is part of Erycidae family (Pyron *et al.*, 2014), and Superfamily Typhlopoidea contains family Typhlopidae with *Xerotyphlops vermicularis* and *Indotyphlops braminus*. For the latter, Wallach (2020) proposed the new genus name *Virgotyphlops* due to its obligate parthenogenetic reproduction, but pending an investigation it is still listed as *Indotyphlops braminus* as in Speybroeck *et al.* (2020).

We have considered the False smooth snake from Lampedusa (Italy) as a separate unit. Based on morphological features (Wade, 2001), most of the recent literature attributes this population to *Macroprotodon cucullatus textilis* (Duméril & Bibron, 1854) (Kreiner, 2007; Capula *et al.*, 2011; Sindaco *et al.*, 2013; Cattaneo, 2015; Di Nicola,

2019). However, genetic studies on mitochondrial markers indicate that this population belongs to a possible specific rank clade that also includes samples from central Tunisia (Faraone et al., 2020). This still unnamed taxon is highly genetically distinct from the other Macroprotodon species present in the European range (M. cucullatus: Kimura 2-parameter distance based on cyt b = 12.9%; M. brevis: Kimura 2parameter distance based on cyt b = 13.4%) (Faraone et al., 2020); furthermore, it is morphologically discriminable from both M. cucullatus and M. brevis (see WADE, 2001). Therefore, in this context we have provisionally named it "Macroprotodon Lampedusa/Tunisia clade".

We have not included in the key the genus *Gloydius* because its presence in Europe (i.e. west side of the Ural river) has not been ascertained yet (see Speybroeck *et al.*, 2020).

Macrovipera schweizeri taxonomic status is still debated: Speybroeck et al. (2020) consider the taxon at a specific level, since the subspecies rank suggested by STÜMPEL & Joger (2009) and Stümpel (2012 - PhD dissertation) on a molecular basis is not supported by data published in peer review literature yet and the relevant Macrovipera sequences are not available in GenBank as well; Freitas et al. (2020) suggest to consider the Milos viper as a subspecies of M. lebetinus, based upon a genetic distance of only 2% (cyt-b) between the two taxa. In this work we maintain the specific status (just like several authors who recently cited the Milos viper in their works -e.g., Cattaneo, 2020; Pizzigalli et al., 2020; Chowdhury et al., 2021; Degen & Brock, 2021; Kontsiotis et al., 2022) according to the most recent checklist by the Taxonomic Committee of the Societas Europaea Herpetologica (Speybroeck *et al.*, 2020).

Some taxa described within the genus *Vipera*, such as *V. lotievi*, *V. magnifica*, *V. nikolskii*, *V. orlovi* and *V. shemakhensis* are not considered valid species according to both Freitas *et al.* (2020) and Speybroeck *et al.* (2020).

At last, also Vipera walser Ghielmi et al., 2016 is currently no longer considered a valid species. Although Freitas et al. (2020) asserted the taxon validity arguing that it has been investigated with an efficient integrative approach (investigating phylogenetic divergence and phenotypic variability), and Seghetti et al. (2020) stated that V. walser is clearly distinct from other Italian vipers based on differences in skull osteology, Speybroeck et al. (2020) consider the acceptance of the new species premature until further nuclear DNA data would confirm the diversity of the walser population. Further nuclear data have been provided by Doniol-Valcroze et al. (2021), showing that V. walser and V. berus marasso are not recovered as distinct lineages due to cyto-nuclear discordance. On the same basis, also SINDACO & RAZZETTI (2021), in the recent checklist of Italian amphibians and reptiles, do not recognize V. walser as a valid species and propose a subspecies rank (*V. berus walser*).

The supporting material file contains the updated checklist of European snakes (Table S1) and the photographic recognition of the species (Fig. S2), numbered and arranged according to the indications of this dichotomous key.

For the nomenclature of the main ce-

phalic scales and for the count of ventral and dorsal scales see the supplementary file in DI NICOLA (2019).

#### RESULTS

### Key to snakes

- **1a.** Worm-like. Dorsal and ventral scales identical, spiny scale on the tip of the tail. Snout rounded in profile, very small eye protected by a semi-transparent shield and visible only as a dark spot......(Typhlopidae) 2
- **1b.** Snake-like. Ventral scales larger than dorsal ones. Eyes well developed and visible......3
- 2a. Very small size (up to 17 cm of total length). Ocular plate divided (eye below the suture of two plates). 20 rows of dorsals at mid-body. Allochthonous, probably native to India and SE Asia. European distribution: Portugal (Madeira), Spain (Canary Islands, Mallorca and one mainland population in Almería Province), Malta and Italy (Sicily chia).....Indotyphlops braminus
- **2b.** Small size (up to 36 cm of total length). Ocular plate entire. Usually 22-24 (rarely 20 or 26) rows of dorsals at midbody. European distribution: Serbia, Montenegro, S Balkans, Bulgaria, several Greek Islands, Turkish Thrace, Cyprus, S European Russia, NE Azerbaijan......

# .....Xerotyphlops vermicularis

**3a.** Subcaudals single (or mostly single). Ventrals wide and covering less than half the width of the belly; remaining belly surface covered with small and smooth identical scales as for the dorsum. Head not distinct from the body. Bulky body. Tail short and blunt. Eyes with vertical pupils. Dorsals of the front half of the

body smooth, keeled towards the end. 41-57 rows of dorsals at midbody.....(Erycidae) 4

- **3b.** Subcaudals paired. Ventrals almost entirely covering the width of the belly. Head more or less distinct from the body. Slenderer body. Longer and pointed tail. Eyes with round or vertical pupils. Less than 30 rows of dorsals at mid-body...........5

- **5a.** Top of the head covered by small scales irregularly arranged or at most three shields symmetrically arranged, surrounded by smaller scales. Anal plate usually entire. Eyes with vertical pupils (slit-like in daylight) and dorsals of the trunk keeled................(Viperidae) 46
- **6a.** Evident concavity on top of the head. Frontal scale at its mid-length clearly narrower than supraoculars. Three contig-

uous scales (two loreals and one preocular) between nasal and eye. Dorsals of the trunk slightly grooved. Typical ridge on top of the eyes consisting of supraocular and the top of the preocular......

## ......Genus Malpolon (Psammophiidae) 7

- **7b.** Dark saddle on the back often present in adult males and even some adult females. Usually 19 dorsal scales at midbody. European distribution: Iberian Peninsula, Balearic islands (recently introduced), S France and NW Italy......

# .....Malpolon monspessulanus

- **8a.** Dorsals clearly keeled, first two rows on the flanks weakly keeled or smooth. One large anterior temporal scale. Usually two or three postoculars. Usually 19 or 21 (rarely 17-23) rows of dorsals at mid-body. Often found close to or in water.................(Natricidae) 9
- **8b.** Dorsals smooth (sometimes barely keeled) or keeled (in this case, only associated with 23, 25 or rarely 27 rows of dorsals at mid-body). Two anterior temporal

scales (occasionally one or three). Usually two postoculars.....(Colubridae) 13

## Key to Natricidae

**9b.** Nostrils pointing upwards. Keeling of dorsals continuous towards the end of the tail. Nape always lacking an evident coloured collar. Usually two preoculars (rarely one or three) and two to four post-oculars. Seven or eight supralabials. Usually 19 or 21 rows of dorsals at mid-body...**12** 

**10a.** Reddish iris. Collar and body markings in juveniles only. Adults uniformly olive-green, brown or grey. European distribution: Iberian Peninsula, SW France (only Aude, where also *N. helvetica* occurs, and Pyrénées-Orientales)......

**10b.** Yellow, grey, whitish or orange iris. Collar and body markings even in

adults. Usually grey body.....11

11b. Collar with larger white/yellowish

## Key to Colubridae

13b. Pupils round, if oval never slit-

shaped. Two contiguous scales between	mented (it can be also entire as in 16a, or
nasal and eye (loreal and preocular)14	short as in 17a). European distribution:
14a. Neck not clearly distinct. Narrow	Italy (only Lampedusa)
crossbands from the back downwards and	Macroprotodon Lampedusa/Tunisia clade
chain-like pattern along the flanks. Scales	18a. Dorsals at mid-body weakly but
smooth and shiny. Juveniles similar to	clearly keeled (keeling decreases as it de-
adults. Usually 21 rows of dorsals at mid-	scends towards the ventral scales). Usually
body. Allochthonous, native to N America.	25 (sometimes 21-27) rows of dorsals at
European distribution: Spain (only Gran	mid-body19
Canaria)Lampropeltis getula	<b>18b.</b> Dorsals smooth or only slightly
<b>14b.</b> Neck usually distinct from the	keeled. Variable number of scale rows at
body. Different pattern15	mid-body <b>24</b>
<b>15a.</b> 6 <sup>th</sup> supralabial in contact or close to	19a. Dark body colouration (often
the parietal. Eight (rarely seven or nine)	black) with narrow light bands. Allochtho-
supralabials and two (rarely one) postocu-	nous, native to NE Asia. European distri-
lars	bution: only NE Netherlands (Drenthe
15b. Supralabials well separated from	Province)Elaphe schrenckii
the parietal18	19b. Different colour and pattern. Out-
<b>16a.</b> Usually 21 (rarely 19-23) rows of	side the Dutch Province of Drenthe20
dorsals at mid-body. Postorbital stripe ex-	20a. Light brown body colouration with
tends from the eye to the corner of the	a dark interconnected H-shaped dorsal
mouth. Dark collar present. Dorsal colour	pattern starting 3-4 head lengths behind
usually greyish and ventral scales charac-	the cranium, fading into a light stripe to-
terized by large dark blotches. European	wards the tail; marked postocular black
distribution: Central and S Iberian Penin-	streak. Allochthonous, native to SE Asia.
sula <i>Macroprotodon brevis</i>	European distribution: only NE Belgium
<b>16b.</b> 19 rows of dorsals at mid-body.	(Limburg Province)Elaphe taeniura
Dark collar fragmented or absent, belly	20b. Different colour and pattern. Out-
immaculate or adorned by small blotch-	side the Belgian Province of Limburg21
es <b>17</b>	21a. Usually rather light brown with
17a. 6th supralabial usually in contact to	four dark stripes along the body (fading
the parietal scale, yellowish-brown ground	towards the tail). Juveniles with a row of
colour, especially along the ventral and	dark, often black bordered, irregular spots
labial scales. Postorbital stripe reduced to a	or bars on the back and one or two series
short spot behind the eye. European distri-	of smaller spots on flanks (sometimes even
bution: Spain (only Mallorca and Menor-	on belly), on grey ground colour.
ca)Macroprotodon cucullatus	Subadults with intermediate colours and
17b. 6th supralabial separated from the	ornamentations between adults and juve-
parietal scale, greyish ground colour	niles. 25 (rarely 23, 26 or 27) rows of dor-
(brownish mainly in the pre-shedding	sals at mid-body. European distribution:
phases). Postorbital stripe mostly frag-	Central and S Italy, from Slovenia along

23a. Males with relatively shorter pileus, lower rostrum, but longer frontal plate and posterior inframaxillary scute. Upper head surface less convex near orbits, prefrontals and internasals. Rostrum less pronounced. Usually 75 subcaudal pairs and one or three loreal scales. Whitish area separating two blotches behind the head. Lateral sides of the head with a dark stripe running from behind the eye towards the corner of the mouth, clearly separated by lighter colour (from the darker head colouration). Spots are usually elongated and yellow or yellowish. European distribution: NE Greece, Turkish Thrace, Bulgaria, Romania, Moldova, S Ukraine, S European Russia, W Kazakhstan...*Elaphe sauromates* 

23b. Males with relatively longer pileus, higher rostrum, but shorter frontal plate and anterior inframaxillary scute. Upper head surface more convex near orbits, prefrontals and internasals. Rostrum more pronounced. Usually 64 subcaudal pairs and one or two loreal scales. Generally darker colouration with dorsal side of the head very dark (sometimes almost black). Lateral sides of the head with a dark stripe running from behind the eve towards the corner of the mouth, less distinguished from head colour. Dorsal body spots are more conspicuous, rounded and typically lined with whitish colour. European distribution: S European Russia, NE Azerbaijan.....Elaphe urartica

**24a.** Nostril in a single nasal scale. Seven supralabials. 15 or 17 rows of dorsals at mid-body. Anatolia and Middle East.......**25** 

25a. 15 rows of dorsals at mid-body...26

**26a.** Dark collar on the neck, consisting in an evident wide band hardly visible from ventral view. On the head of the juveniles, three dark transverse and independent bands of variable width, which can form a crown-like spot. Body colour usually uniform with lighter scales in the

centre. Seven to nine supralabials, small	<b>30b.</b> Different pattern, colouration and
rostral scale non wedged between the in-	body shape32
ternasals. European distribution: S Europe-	31a. Neck with a dark, white bordered
an Russia, NE Azerbaijan	collar. Eyes with dark anterior and posteri-
Eirenis collaris	or borders. Spots irregularly decreasing in
26b. Usually black head with white su-	size along the body and more separated
pralabial. Unpatterned back with orange,	from each other. European distribution: E
grey or light brown colour. Six supralabi-	Bulgaria, Turkish Thrace
als, prominent rostral scales wedged be-	Platyceps collaris
tween the internasals. European distribu-	31b. Neck with two dark, white bor-
tion: Cyprus	dered spots on sides, sometimes joined in
Rhynchocalamus melanocephalus	a collar. Eyes with white anterior and pos-
27a. Head pattern: dorsals without dark	terior borders. Spots gradually decreasing
centre and parietal band limited to the pa-	in size along the body and less separated
rietal scales. Three to five scales from the	from each other. European distribution:
anterior beginning of the paravertebral	from Croatia along the E Adriatic Coast to
scale row to the posterior border of the	S Balkans, Serbia, Bulgaria, several Greek
collar. European distribution: Cyprus	islands, Cyprus, S European Russia, NE
Eirenis levantinus	Azerbaijan <i>Platyceps najadum</i>
27b. Head pattern: interocular and pari-	32a. Seven (rarely eight) supralabials,
etal band scythe-like and parietal band	with 3 <sup>rd</sup> and 4 <sup>th</sup> in contact with the eye.
sometimes joined to the interocular in the	Dark band from nostrils to the sides of the
median line. One or two scales from the	neck across the eyes and sometimes a blur-
anterior beginning of the paravertebral	ry mark from eye to eye. Dark ventrals.
scale row to the posterior border of the	European distribution: all Europe except
collar. European distribution: some Greek	Iceland, Ireland, Central and N Great Brit-
islands, Turkey (Istanbul area), S European	ain and N Scandinavia; not uniform distri-
Russia, NE AzerbaijanEirenis modestus	bution in E EuropeCoronella austriaca
28a. 17 or 19 rows of dorsals at mid-	32b. Eight supralabials. Different
body <b>29</b>	pattern33
28b. More than 19 rows of dorsals at	33a. Greenish-yellow ground colour,
mid-body37	often with predominant black or dark
29a. 17 rows of dorsals at mid-body.	green crossbars on foreparts. Rest of the
European distribution: endemic to Cy-	body with yellowish longitudinal streaks.
prusHierophis cypriensis	Melanism prevailing in the eastern and
<b>29b.</b> 19 rows of dorsals at mid-body <b>30</b>	southern part of its range. 187-227 ventrals.
30a. Neck with dark spots or a dark	European distribution: Pyrenees, France, S
collar. Body with rather "double" coloura-	Switzerland, Italy, W Slovenia, NW Croa-
tion, first greyish and then beige-brown till	tia, MaltaHierophis viridiflavus
the tail tip. Dorsals with single apical pits.	33b. Different colour34
Extremely slender body31	34a. Dark blotches and often small light

spots and streaks on foreparts. Dorsals	of dorsals at mid-body. European distribu-
always narrowly pale edged. Melanism	tion: Iberian Peninsula, S France, Centra
absent. 167–186 ventrals. European distri-	and N ItalyCoronella girondica
bution: from Slovenia along the E Adriatic	38b. Head and belly pattern different
Coast to Greece, Serbia, several Greek is-	Usually 23 (rarely 21) or more rows of dor
landsHierophis gemonensis	sals at mid-body39
34b. Rather uniform colouration. Dor-	39a. Rostral shield large and pointed
sals usually not narrowly pale edged. Mel-	between the internasals. Dorsum with two
anism frequent. More than 185 ventrals.	evident longitudinal dark stripes on grey
Large sized (up to 250 cm)35	or beige ground colouration. Dark H
<b>35a.</b> Black dorsum. Top of the head	shaped blotches on juveniles' back, ever
black or slightly lighter, with brown	connected in a ladder design. 27 (rarely 25
shades. European distribution: some S	or 29) rows of dorsals at mid-body. 201
Greek islands, Cyprus	220 ventrals. European distribution: Iberi
Dolichophis jugularis	an Peninsula, Balearic islands (recently
35b. Not black dorsum36	introduced), S FranceZamenis scalaris
36a. Dorsum greyish-brown to oliva-	<b>39b.</b> Rostral shield not pointed between
ceous (greyish-brown in preserved speci-	the internasals. Different pattern, with o
mens), sometimes copper head. Light lon-	without stripes (which are not so evi
gitudinal central streak on dorsal scales.	dent)40
Belly white to yellow. European distribu-	<b>40a.</b> Ventrals not keeled. Reddish of
tion: Balkans, Hungary, Moldova, Ukraine,	brown black-bordered blotches
S European Russia, W Kazakh-	(sometimes connected in longitudina
stanDolichophis caspius	lines) on the back41
<b>36b.</b> Dorsum orange to bright brick red	<b>40b.</b> Ventrals strongly keeled. Juveniles
(light brown to reddish brown in pre-	with spotted pattern and two dark streaks
served specimens). Dark longitudinal cen-	one from the eye to the corner of the
tral streak of variable intensity on dorsal	mouth and one under the eye, that both
scales. Belly yellow to orange (ochraceous	faint with ageing. 23 (rarely 21) rows of
in preserved specimens). European distri-	dorsals at mid-body42
bution: S European Russia, NE Azerbai-	41a. Ochre or light brown spots with
jan <i>Dolichophis schmidti</i>	black borders on the back (two rows sepa
37a. One preocular, pre-subocular ab-	rated by a vertebral light line in ssp. ho
sent38	henackeri or one row of connected spots in
37b. One or more pre-suboculars pre-	ssp. taurica). Ochre iris. 23 (rarely 25) rows
sent43	of dorsals at mid-body. 203-230 ventrals
38a. Dark band from eyes to the sides	European distribution: S European Russia
of the neck and from eye to eye. Another	(close to Caucasus)Zamenis hohenackern
dark streak under the eye. Belly chequered	<b>41b.</b> Orange to brown spots with black
or striped. 8 supralabials, with 4th and 5th	borders on the back or two dorsal longitu
in contact with the eye. 21 (rarely 23) rows	dinal stripes with black borders (at least or
	i carried the second control of

**42a.** Adults often with four longitudinal dark stripes but blurry and not always clearly distinct from ground colouration. Nape without blotches. Belly greyish. Orange to red iris. European distribution: endemic to S Italy and Sicily......

.....Zamenis lineatus

**42b.** Adults usually with grey-brown, greenish or blackish uniform colouration, often with small white dots, something with four faint longitudinal dark stripes. Nape with light blotches. Belly yellowish. Ochre iris. European distribution: NE Spain, mainland France, Great Britain (recently introduced), N and Central Italy, Central Europe, Balkans, Moldova, S and W Ukraine, S European Russia. In Apulia (SE Italy) there are individuals genetically attributable to this species but with an intermediate morphology between *Z. longissimus* and *Z. lineatus*......

## .....Zamenis longissimus

- **44a.** Dorsal roundish spots usually separated. 23 rows of dorsals at mid-body. 82-107 subcaudals in males, 82-107 in females. European distribution: several Greek is-

**45a.** Dorsal colour with widely separated spots. Head pattern with more or less distinct transverse bar or dark blotch on nape. Usually one supralabial touching the eye not necessarily on both sides of the head. 23-25 rows of dorsals at mid-body. European distribution: Malta......

......Hemorrhois algirus

45b. Dorsal colour with large dark polygonal spots (with darker borders) clearly distinct from ground colouration. Typical horseshoe marking on head. No supralability als touching the eye due to the presence of a series of small suboculars. 27-29 (rarely 25) rows of dorsals at mid-body. European distribution: Iberian Peninsula, Balearic island (recently introduced), Italy (Sardinia and Pantelleria).......

......Hemorrhois hippocrepis

### Key to Viperidae

- **47a.** Supraoculars large. European distribution: Greece (eastern Thrace), Turkish Thrace and several Greek and Turkish islands from Dodecanese Archipela-

go......Montivipera xanthina

distribution: endemic to S Albania and

47b. Supraoculars divided48	GreeceVipera graeca
48a. 23 (rarely 19-25) rows of dorsals at	<b>52b.</b> 21 rows of dorsals at mid-body.
mid-body. Dorsal pattern consisting of two	Larger body (50/60 cm, up to 65 cm). Su-
series of transverse alternate bars, often	pralabials rather similar53
light and not so evident and joined form-	53a. 21 rows of dorsals at mid-body,
ing a zigzag band. European distribution:	reduced to 19 rows on the second part of
endemic to Greece (W Cyclades islands of	the body. 135-150 ventrals. European dis-
Kimolos, Milos, Polýaigos and Sifnos)	tribution: Ukraine, Russia, W Kazakh-
Macrovipera schweizeri	stanVipera renardi
48b. 25 (rarely 23) or more rows of dor-	53b. 21 rows of dorsals on the first part
sals at mid-body. Dorsal pattern consisting	of the body, reduced to 19 rows before
of two series of transverse alternate dark	reaching the mid-body. 133-135 ventrals
bars, often joined forming a zigzag band.	(or more)54
European distribution: S European Russia,	54a. Usually one apical in contact with
NE Azerbaijan, Cyprus	the rostral shield. Upper side of the naso-
Macrovipera lebetinus	rostral shield straight. Upper preocular in
49a. Snout rounded. Frontal and parie-	contact with the nasal. Head with large
tal scales mostly present50	shields. Usually less than 13 intercanthals
<b>49b.</b> Snout upturned or horned. Frontal	+ inter-supraoculars. No yellow, orange or
and parietal mostly replaced by smaller	red in dorsal background colour. Absent
scales56	from Caucasus territory. European distri-
<b>50a.</b> Nostril in the centre of the nasal <b>51</b>	bution: SE France, Central Italy, Austria,
<b>50b.</b> Nostril close to the lower edge of a	Hungary, Balkans, Moldova
rather large nasal shield52	Vipera ursinii
<b>51a.</b> Head clearly distinct from the	<b>54b.</b> Usually two apicals in contact with
body. Snout just slightly sharp. Parietals	the rostral shield. Upper side of the naso-
and frontal variably fragmented. European	rostral shield rounded. Upper preocular in
distribution: N Iberian Peninsula, SW	contact with or separated from the nasal.
FranceVipera seoanei	Dorsal background colour usually yellow,
<b>51b.</b> Head moderately distinct from the	orange or red. Present on Caucasus territo-
body. Snout rounded. Parietals and frontal	ry55
usually entire. European distribution: al-	<b>55a.</b> Head narrow, barely distinct from
most all of Europe (often restricted to	the body and flat or convex above. Head
higher elevations in W and C of the conti-	less wide than the distance from snout tip
nent), except Caucasus, Iberian Peninsula,	to the angle of the mouth. Dorsal pattern
Iceland and IrelandVipera berus	different. There are totally melanistic indi-
<b>52a.</b> Sometimes just 17 rows of dorsals	viduals, without light spots. Rostral shield
at mid-body. Shorter body (up to 35 cm,	is usually in contact with one apical scale.
females rarely up to 45). 2 <sup>nd</sup> , 3 <sup>rd</sup> and 4 <sup>th</sup> su-	European distribution: E Caucasus Moun-
pralabials wide under the eye. European	tainsVipera dinniki

**55b.** Head wide, clearly distinct from the body. Head as wide as the distance from snout tip to the angle of the mouth. Rostral shield is usually in contact with two apical scales. Dark or black ground colour (with a yellow to red pattern that always persists in labials of melanistic specimens). European distribution: W Caucasus Mountains......Vipera kaznakovi

**57a.** Rostral scale scarcely reaching an evident horn, usually covered with 5-20 scales. 21 rows of dorsals at mid-body (occasionally 20-23). European distribution: NE Italy, S Austria, Balkans, several Greek islands, Turkish Thrace......

.....Vipera ammodytes

#### Discussion

Following the latest taxonomic updates and the recent species considered part of the European herpetofauna, the checklist of European snakes currently has 57 species (see Table S1) as well as one taxon that needs to be better defined (Macroprotodon Lampedusa/Tunisia clade). Four out of 57 of these species are allochthonous to our range, thus adding one more non-native species (Elaphe taeniura) to the three (E. schrenckii, Indotyphlops braminus and Lampropeltis getula) already listed in the checklist by Speybroeck et al. (2020). One species (Rhynchocalamus melanocephalus), recently found in Cyprus, may be attributed to human-mediated dispersion or natural colonization, since a Cypriot specimen showed genetic similarity to another one from northern Israel; further investigations will be needed to clarify the origins of the specimens from Cyprus (see Tamar et al., 2020).

The dichotomous key provided here allows the identification by using morphological characters (especially pholidosis and colouring) together with the area of discovery. This key is therefore to be considered a field instrument, but it can be useful even with museum specimens if the samples are arranged with the place of origin. Moreover, this work can make the identification of snake species for the citizen science more accessible, thus enhancing data collection and assist species conservation. Another silver lining is dealing with monitoring the diffusion of allochthonous species, making easier to detect them in new territories and limit their expansion.

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